IMPORTANT: This syllabus form should be submitted to OAA (<u>gsbs_academic_affairs@uth.tmc.edu</u>) a week before the start of each semester.

NOTE to STUDENTS: If you need any accommodations related to attending/enrolling in this course, please contact one of the Graduate School's 504 Coordinators, Cheryl Spitzenberger or Natalie Sirisaengtaksin. We ask that you notify GSBS in advance (preferably at least 3 days before the start of the semester) so we can make appropriate arrangements.

Term and Year: Spring 2023	Program Required Course: No
Course Number and Course Title:	Approval Code: No
Imaging Cells and Molecules	Audit Permitted: Yes
Credit Hours: 1	Classes Begin: 1/09/2023
Meeting Location: McGovern Medical School	Classes End: 4/10/2023
Building/Room#: MSB 1.180	Final Exam Week: 4/17/23
WebEx/Zoom Link: N/A	

Class Meeting Schedule

Day	Time 1-2pm (Lectures), 2-4pm (Labs)	
Monday		
Course Director	Instructor/s	
Name and Degree: Bo Hu, Ph.D. Title: Assistant Professor	1. Bo Hu, Ph.D. Institution: UTH Email Address: <u>Bo.Hu@uth.tmc.edu</u>	
Institution: UTH Email Address: <u>Bo.Hu@uth.tmc.edu</u>	2. William Margolin, Ph.D. Institution: UTH Email Address: <u>William.Margolin@uth.tmc.edu</u>	
Contact Number: 713-500-5434 Course Co-Director/s: Name and Degree: William Margolin, Ph.D.	3. Kevin Morano, Ph.D. Institution: <i>UTH</i> Email Address : <u>Kevin.A.Morano@uth.tmc.edu</u>	
Title: Professor Department: Microbiology & Molecular Genetics Institution: UTH	4. Irina I. Serysheva, Ph.D. Institution: UTH Email Address: Irina.I.Serysheva@uth.tmc.edu	
Email Address: <u>William.Margolin@uth.tmc.edu</u> Contact Number: 713-500-5452	5. Kuang-Lei Tsai, Ph.D. Institution: UTH Email Address: <u>Kuang-Lei.Tsai@uth.tmc.edu</u>	

NOTE: Office hours are available by request. Please email me to arrange a time to meet.	6. Travis I. Moore, Ph.D. Institution: <i>UTH</i> Email Address: <u>Travis.I.Moore@uth.tmc.edu</u>
Teaching Assistant: (if any)	
N/A Name and Email Address	7. Heidi B. Kaplan, Ph.D. Institution: UTH Email Address: <u>Heidi.B.Kaplan@uth.tmc.edu</u>
	8. Todd Cameron, Ph.D. Institution: UTH Email Address: <u>Todd.Cameron@uth.tmc.edu</u>

Course Description:

The Fluorescence and Electron Microscopy course consists of lectures and labs on Monday afternoons during the Spring semester. The most important and valuable feature of this course is that provides hands-on experience with fluorescence microscopy as well as transmission electron microscopy and cryo-EM, taught by experts in various specialties.

Textbook/Supplemental Reading Materials (if any)

• N/A

Course Objective/s:

Upon successful completion of this course, students will ...

Specific Learning Objectives:

- 1. Learn about the optical principles underlying light microscopy and basic microscope hardware employed for this type of imaging.
- 2. Understand the basis of cutting-edge fluorescence microscopy methods for imaging of cells and subcellular organelles, with an emphasis on microbial cells.
- 3. Learn how to acquire, process and analyze images of cells and subcellular localization patterns obtained from light and fluorescence microscopy.
- 4. Understand the principles and basics of electron microscopy, and learn about sample preparation and operation of an electron microscope.
- 5. Understand the basis of cryo-electron microscopy, and learn about cryo sample preparation and basics of 3D reconstruction from 2D tilt serials.

Student Responsibilities and Expectations:

Students enrolled in this course will be expected to perform the following activities each week.

- 1. Read and review (study) assigned reading material (reviews and research literature) prior to class.
- 2. Attend and participate at the lab session.
- 3. Participate in and contribute to course discussions during lecture.
- 4. Prepare for homework based on lecture and lab practice.

Students are expected to complete all assigned reading material (reviews and research literature) prior to class. While you may work and discuss all course materials and assignments in groups, all writing assignments must be your own. Plagiarism and failure to properly cite scientific literature and other sources will not be tolerated and are grounds for dismissal from the course and further GSBS disciplinary action. Cheating or engaging in unethical behavior during examinations (quizzes and final) will be grounds for dismissal from the course without credit and further GSBS disciplinary action.

Grading System: Pass/Fail

Percentage	Description	
Homework (30%)	Students will write Materials & Methods and Results Sections accompanied with a publishable multi-panel figure and a figure legend corresponding to data obtained from each section, i.e. fluorescence microscopy (Margolin, Cameron, Kaplan) for Assignment 1, electron microcopy (Tsai) for Assignment 2, and cryo-EM (Hu and Serysheva) for Assignment 3.	
Quiz (%)	N/A	
Midterm Exams (%)	N/A	
Final Exam (%)	N/A	
Participation and/or Attendance (70%)	Attendance (20%), Lab participation (50%)	

Student Assessment and Grading Criteria : (May include the following:)

CLASS SCHEDULE

	Duration		
	(Hour(s)		
Date	lecturer)	Lecture Tonic	lecturer/s
Dute	lecturery		
<u>1/09/2023</u>		Lecture: Principles of Light Microscopy	Morano
		Lecture: Hardware and Equipment	
		Lab: Preparation of samples, fluorescence	
<u>1/23/23</u>		microscopy	Morano/Cameron/Margolin
		Lecture: Whole Cell Imaging	
		Lab: More Fluorescence Microscopy and Confocal	
<u>1/30/23</u>		Microscopy	Margolin/ Kaplan
		Lecture: Image Post-Processing	
<u>2/06/23</u>		Lab: Image Processing and Analysis	Margolin/ Cameron
		Lecture: Advanced Fluorescence Microscopy	
		Technologies	
		Lab: Super-Resolution and TIRF Fluorescence	
<u>2/13/23</u>		Microscopy	Margolin/ Moore
		Lecture: Introduction to Electron Microscopy	
<u>2/20/23</u>		Lab: Operational principles of TEM	Tsai
<u>2/27/23</u>		Lab: Specimen Preparation for TEM	Tsai
<u>3/06/23</u>		Lab: TEM – Operation and Data Collection	Tsai
		Lecture: Cryo-electron microscopy	
<u>3/20/23</u>		Lab: Preparation of biological samples for cryo-EM	Hu
		Lecture: Cryo-EM – Operation and Data Collection	
<u>3/27/23</u>		Lab: Cryo-EM – Operation and Data Collection	Hu
<u>4/03/23</u>		Lab: Cryo-EM – Image Processing	Hu
		Lecture (2 hours): Electron Cryomicroscopy	
4/10/23		Lab: Specimen preparation for cryo-EM	Serysheva

BH/jal